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ARTICLE Performance and Fatty Acids Composition of Oreochromis Niloticus Fed on Maggots

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ABSTRACT

This study was conducted to assess **improved performance and fatty acids composition of** *Oreochromis niloticus*. Different substrate was used to culture and analyze the effect. Growth performance of *O. niloticus* fed on HFM meal and enhancement of fatty acids in the produced *O. niloticus* was conducted. Results showed that fish fed on HFMEuch achieved high performance with higher amount of ω -3FAs levels accumulation. *Eucheuma* species can be used to culture HFM as alternative non-competitive feed ingredient to improve performance and composition of ω -3FAs in cultured *O. niloticus*.

1. Introduction

Tanzania faces high demand of quality protein for human consumption and also for animal feeds. Approximately 85% of fish production is used for consumption while the rest is for non-human uses ^[2].

High costs of important ingredients such as fish meal which are used in making fish feeds is a limiting factor in growth of aquaculture industry ^{[3], [4]}. However, due to high demand for human consumption, this leads to requiring alternative non-competitive protein source.

Fish are good source of fatty acids which is important for different body physiological activities. However, freshwater fish, including tilapias which are highly depended fish in Tanzania. Previous studies show that supplementation of tilapia with feed having high concentration of ω -3 fatty acids can increase the amount of these fatty acids in the muscle tissues of fish ^[8].

Lemna has high composition of fatty acids and can

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enhance fatty acids in fish dietary ^[11]. *Eucheuma* provides provide good lipid and quality protein however of its un-palatability nature ^[12]. Housefly maggots accumulates nutrients from substrate and transfer to fish when used as feed ingredient ^[15].

This study evaluated improvement of performance and fatty acids composition of *Oreochromis niloticus*.

2. Materials and Methods

Two species of aquatic macrophyte *Lemna* and *Eucheuma spp* were collected from Lake Victoria and from Indian Ocean, respectively. *Lemna* sp was sorted to remove unwanted materials and debris, and then were fermented for three days to acquire offal odor to attract housefly and make them useful for maggot's culture. *Eucheuma sp* was cleaned then were fermented for three days ready for maggot's culture at experimental site.

Culturing was done indoors for each of the substrates as described by ^[16] Devic (2014) and ^[17]. The mature maggots were harvested according to ^[18]. The samples for formulate feed including maggot meal were taken to Laboratory for proximate analysis as shown in table 1 below.

2.1 Determination of fish growth and feed utilization

The body weights of fish from each replicate were recorded in bulky and finally mean weights was calculated.

2.2 Lipid extraction and fatty acids analysis

Frozen fish sample were taken to the Zoology laboratory for lipid extraction and fatty acids analysis

2.3 Lipid extraction

Extraction of lipid was done by using Folch Methods

3. RESULTS

There was no significant difference from all substrates (P>0.05) as shown in table 2 below.

There was no feed related mortality observed during the entire period of the experiment.

	SBM	FM,	HFMChick	HFMLemn	HFMEuch
SBM	51.49	43.90	12.72	10.80	7.50
FM	0.00	5.00	0.00	0.00	0.00
MM	37.70	40.33	40.15	43.33	44.75
HMM	0.00	0.00	35.00	35.00	35.00
CRM	2.49	2.48	3.90	2.53	4.33
CGM	0.00	0.00	0.00	0.00	0.00
SFO	4.32	4.29	0.46	0.47	0.65
Premix	1.00	1.00	1.00	1.00	1.00
Meth	1.00	1.00	1.00	1.00	1.00
Lysine	1.00	1.00	1.00	1.00	1.00
МСР	1.00	1.00	1.00	1.00	1.00
Total	100.00	100.00	100.00	100.00	100.00

Table 1: Diet formulation and grouping

Table 2: Chemical composition of formulated diets

Inquedient (0/)	Diets							
Ingredient (%)	SBM	FM	HFMChick	HFMLemn	HFMEuch			
Dry matter	90.75	90.89	88.42	91.86	91.40			
Crude protein	41.73	46.01	40.66	46.03	50.00			
Ether extract	18.98	19.20	20.00	19.07	20.40			
Crude fibre	1.45	1.90	1.22	0.86	1.08			
Ash	7.57	7.66	7.31	7.69	7.73			

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	Diets					
Parameter	SBM	FM	HFMChick	HFMLemn	HFMEuch	P-value
INBWT (g)	$2.41\pm0.17^{\rm a}$	2.48 ± 0.11^{a}	$2.54\pm0.12^{\text{a}}$	$2.32\pm0.08^{\rm a}$	$2.52\pm0.04^{\text{a}}$	0.344
FBWT (g)	$7.71.\pm0.07^{a}$	$8.19\pm0.19^{\rm a}$	8.07 ± 0.19^{a}	$7.81\pm0.37^{\rm a}$	$8.33\pm0.20^{\text{a}}$	0.048
BWTG (g)	$5.01\pm0.27^{\rm a}$	$5.65\pm0.20^{\rm a}$	$5.29\pm0.34^{\rm a}$	$5.01\pm0.27^{\rm a}$	$5.59\pm0.08^{\rm a}$	0.161
ADG(g/day)	$0.100{\pm}0.005^{a}$	0.101 ± 0.001^{a}	$0.091{\pm}0.006^{b}$	$0.100{\pm}0.005^{a}$	$0.100{\pm}0.041^{a}$	0.161
SGR (%day)	$2.00\pm0.15^{\text{b}}$	2.16 ± 0.09^{a}	$2.11\pm0.049^{\text{a}}$	$1.88\pm0.06^{\text{b}}$	$2.18\pm0.06^{\text{b}}$	0.711
FI (g/fish/day)	0.22±0.01 ^b	0.28 ± 0.01^{a}	$0.26{\pm}0.01^{ab}$	$0.24{\pm}0.01^{b}$	0.30 ± 0.01^{ab}	0.045
FCR	$2.24{\pm}~0.07^{abc}$	$2.05\pm0.01^{\text{b}}$	$2.66\pm0.16^{\rm dc}$	$2.47\pm0.01^{\circ}$	$1.85\pm0.04^{\rm a}$	0.073
PER	$1.35\pm0.01^{\text{b}}$	$1.64\pm0.09^{\text{ad}}$	$1.49\pm0.05^{\rm b}$	$1.26 \pm 0.08^{\circ}$	$1.81\pm0.03^{\text{a}}$	0.511
SR (%)	$88.1\pm8.6^{\text{b}}$	$97.6\pm2.4^{\text{a}}$	$95.2\pm4.8^{\rm a}$	$97.6\pm2.4^{\rm a}$	$95.2\pm2.1^{\text{a}}$	0.559
			Table 4:			
Diets						
Parameters	(D1)	(D2)	(D3)	(D4)	(D5)	P value
ΣPUFAS	$1.99\pm0.01^{\text{a}}$	$2.84 \pm 0.10^{\text{b}}$	$0.88\pm0.12^{\rm a}$	$4.81\pm0.05^{\circ}$	$9.52\pm0.82^{\rm c}$	0.0001
Σω-3 PUFAs	$1.54\pm0.06^{\rm a}$	2.83 ± 0.16^{b}	$0.69\pm0.29^{\rm a}$	$2.73\pm0.38^{\text{b}}$	$4.07\pm0.91^{\circ}$	0.0003
Σω-6 PUFAs	0.43 ± 0.05^{a}	0.33 ± 0.0^{a}	0.13 ± 0.04^{a}	1.09 ± 0.0286^{a}	$4.54 \pm 0.37^{\circ}$	< 0.0001

Table 3: Growth performance and nutrient utilization of Nile tilapia fed different diets (mean±SE)

4. Discussion

The findings from this study showed that fish fed on HFMEuch diet the performance were superior compared to fish fed on other diets. Higher growth performance of *O. niloticus* fed on HFMEuch diets reflects palatability and high protein content of maggots cultured in *Eucheuma* species of the marine macrophyte.

FCR obtained from fish fed HFMEuch was the lowest compared to other HFM which implies higher weight gain obtained from the feed. ^[35]. Similar findings were reported previously ^{[14], [31]} and ^[36] whose values ranged from 3.13 to 5.07.

To conclude, housefly maggots cultured in aquatic macrophyte (seaweed and duckweed) are cheap and non-competitive, and fish fed this maggot diet grow fast compared to other diet.

References

- FAO, (2014). Food and Agriculture Organization of the United States. The state of World Fisheries and Aquaculture: Opportunities and challenges. *Food* and Agriculture Oraganization of the United Nations, Rome, Italy, 243 pp.
- [2] Barbaroux, G, Bizzarri, M. R, Hasan, L, Miuccio, J, Saha, J, Sanders, J. S. and Jerome, V. A. (2012). The State of World Fisheries and Aquaculture. Food and Agriculture Organization of the United Nations, Rome, Italy. 230 pp.
- [3] Huntington, T. C. and Hasan, M. R. (2009). Fish as

feed inputs for aquaculture: Practices, sustainability and implications: A global synthesis. In; M. R. Hasan and M. Halwart (eds). Fish as feed inputs for aquaculture: practices, sustainability and implications. Food and Agriculture Organization of the United Nations, Rome, Italy 518(2): 1-61.

- [4] Dedeke, G. A., Owa, S. O., Olurin, K. B., Akinfe, A. O. and Awotedu, O. O. (2013). Partial replacement of fish meal by earthworm meal (*Libyodrilus violaceus*) in diets for African catfish, *Clarias gariepinus*. *International Journal of Fisheries and Aquaculture* 5(9): 229–233.
- [5] Salah, H. H. E. (2020). Review on Using of Housefly Maggots (*Musca domestica*) in Fish Diets. *Journal of Zoological Research* 2(4): 39 – 46.
- [6] Mohanta, K. N., Sankaran, S. and Veeratayya, S. K. (2013). Evaluation of Different Animal Protein Sources in Formulating the Diets for Blue Gourami, *Trichogaster trichopterus* Fingerlings. *Journal of Aquaculture Research and Development* 4(2): 7-15.
- [7] Silva, B. C. E., dos Santos, H. M. C., Montanher, P. F., Boeing, J. S., Almeida, V. D. C. and Visentainer, J. V. (2014). Incorporation of Omega-3 Fatty Acids in Nile Tilapia (*Oreochromisniloticus*) Fed Chia (*Salvia hispanica* L.) Bran. *Journal of the American Oil Chemists' Society* 91(3): 429-437.
- [8] Tiffany, D., Kumar, H., du Toit, E., Kulkarni, A., Aakko, J., Linderborg, K. M., Zhang, Y., Nicol, M. P., Isolauri, E., Yang, B. and Collado, M. C. (2016). Distinct patterns in human milk microbiota and fatty

acid profiles across specific geographic locations. *Front Microbiology* 41: 508–518.

- [9] Whelan, J. (2009). Fishy business: Aquaculture, omega-3 fats and health. Presented at Aquaculture America 2009, Seattle, WA.WWF-US in 2004. Auburn University, Alabama 36831. [https://www. ncbi.nlm.nih.gov/pubmed/ 22233511] site visited on 12/06/2018.
- [10] Landolt, E. and Kandeler, R. (1987). The family of Lemnaceae- A monographic study: Phytochemistry, physiology, application and bibliography, vol. 4. Veroffentlichungen des Geobotanisches Institut der ETH, Stiftung Ruebel, Zurich.
- [11] Leng, R. A. (1999). Duckweed: A Tiny Aquatic Plant with Enormous Potential for Agriculture and Environment. Animal Production and Health Division, University of Tropical Agriculture Foundation, Phnom Penh (Combodia). FAO Rome (Italy). 108pp.
- [12] Norziah, M. H. and Ching, C. Y. (2000). Nutritional composition of edible seaweed, *Gracilaria channgi*, an edible species of nori from Nova Scotia. *Food Chemistry* 68: 69-76.
- [13] Chaturvedi, K. M. M., Langote, D. S. and Asolekar, R. S. (2003). Duckweedfed fisheries for treatment of low strength community waste water. WWWTM Newsletter-Asian Institute of Technology. India. [https://www.researchgate.net/.../ 319059154_Use _ of_duckweed_ Lemna_ minor_as_a_pr...] site visited on 14/09/2018.
- [14] Jabir, M. D. A., Razak, S. A. and Vikineswary, S. (2012). Nutritive potential and utilization of super worm (*Zophobas morio*) meal in the diet of Nile tilapia (*Oreochromis niloticus*) juvenile. *African Journal* of *Biotechnology* 11(24): 6592–6598.
- [15] Negesse, T., Makkar, H. P. S. and Becker, K. (2009). Nutritive value of some nonconventional feed resources of Ethiopia determined by chemical analyses and an in vitro gas method. *Food Science and Technology* 5(3): 204-217.
- [16] Devic, M. (2014). Cell Culture Basics. Invitrogen, Gibco. 62pp.
- [17] Nzamujo, O. P. (2001). Techniques for maggot production: The Shonghai experience. [http://www.ias. unu.edu/proceedings/icibs/ibs/shonghai] site visited on 05/02/2018.
- [18] Sogbesan, A. O., Ajuonu, N., Musa, B. O. and Adewole, A. M. (2006). Harvesting Techniques and Evaluation of Maggot Meal as Animal Dietary Protein Sources for "*Heteroclarias*" in Outdoor Concrete Tanks. *World Journal of Agricultural Sciences* 2(4): 394–402.

- [19] AOAC (2005). Official Methods of Analysis of Association of Official Analytical Chemists. 18th Edition, Washington, DC. [https://www.ncbi.nlm.nih.gov] site visited on 15/09/2022.
- [20] Olvera-Novoa, M. A., Campos, S. G., Sabido, M. G. and Palacios, C. A. M. (1990). The use of alfalfa leaf protein concentrates as a protein source in diets for tilapia (*Oreochromis mossambicus*). Aquaculture 90(3-4): 291-302.
- [21] Folch, J., Lees, M. and Stanley, G. H. S. (1957). A simple method for the isolation and purification of total lipides from animal tissues. *Journal of Biological Chemistry* 226(1): 497–500.
- [22] Dytham, C. (2013). Choosing and Using Statistics: A Biologist Guide. [https://www.researchgate. net/.../248812045_Choosing_and_Using_Statistics] site visited on 12/03/2019.
- [23] Steel, R. G. D. and Torrie, J. H. (1980). Principles and Procedures of Statistics. A biometrical Approach. 2nd edition. McGraw-Hill, New York, USA, pp. 20-90.
- [24] Odesanya, B. O., Ajayi, S. O., Agbaogun, B. K. O. and Okuneye, B. (2011). Comparative Evaluation of Nutritive Value of Maggots. *International Journal of Scientific and Engineering Research* 2 1-9.
- [25] Adeniji, A. A. (2007). Effect of Replacing Groundnut Cake with Maggot Meal in the Diet of Broilers. *International Journal of Poultry Science* 6(11): 822– 825.
- [26] Nzamujo, O. P. (1999). Technique for maggot production. The Songhai experience. [http://ias.unu.edu/ en/proceedings/icibs/ibs/songhai/index.htm] site visited on 05/03/2018.
- [27] Agbeko, E., Sakyi, P., Obeng, A. K. and Quainoo, A. K. (2014). A sustainable production of Maggots (squatts) as live food for Nile tilapia (*Oreochromis* niloticus). International Journal of Multidisciplinary Research and Development 1(1): 51–55.
- [28] Ajonina, A. S. and Nyambi, R. E. (2013). Original Research Article Evaluation of growth response of *Clarias gariepinus* fingerling fed dried maggot as protein source. *International Journal of Current Microbiology Application Science* 2(5): 123-129.
- [29] Patricia, L. S. and Salas, C. (2007). House fly (Musca domestica L.) (Diptera : muscidae) development in different types of manure. Chilean Journal of Agricultural Research 68: 192–197.
- [30] Ogunji, J. O., Kloas, W., Wirth, M., Schulz, C. and Rennert, B. (2006). Housefly Maggot Meal (*Mag-meal*): An Emerging Substitute of Fishmeal in Tilapia Diets. In: *Proceedings of International Agricultural*

Research for Development. 7 pp.

- [31] Omoyinmi, G. A. K. and Olaoye, O. J. (2012). Growth Performance of Nile Tilapia, *Oreochromis niloticus* Fed Diets Containing Different Sources of Animal Protein. *Libyan Agriculture Research Center Journal International* 3(1): 18–23.
- [32] Okah, U. and Onwujiariri, E. B. (2012). Performance of finisher broiler chickens fed maggot meal as a replacement for fish meal. *Journal of Agricultural Technology* 8(2): 471–477.
- [33] Ogunji, J., Toor, R. S., Schulz, C. and Kloas, W. (2008). Growth Performance, Nutrient Utilization of Nile Tilapia Oreochromis niloticus Fed Housefly Maggot Meal (Magmeal) Diets. Turkish Journal of Fisheries and Aquatic Sciences 8: 141–147.
- [34] Makkar, H. P., Tran, G., Heuzé, V. and Ankers, P. (2014). State-of-the-art on use of insects as animal feed. *Animal Feed Science and Technology* 197: 1-33.
- [35] Idowu, A. B., Amusan, A. A. S. and Oyediran, A. G. (2003). The response of *Clarias gariepinus* fingerlings (Burchell 1822) to the diet containing Housefly maggot (*Musca domestica*) (L). *Nigerian Journal of Animal Production* 30(1): 139-144.
- [36] Mekhamar, M., Ali Ahmed, E., Gadel-Rab, I. G. and Osman, A. (2015). Evaluation of Growth Performance of Nile Tilapia *Oreochromis niloticus* fed *Piophila casei* Maggot Meal (Magmeal) Diets. *American Journal of Life Sciences* 3: 1-12.
- [37] Olaniyi, C. O. and Salau, B. R. (2013). Utilization of maggot meal in the nutrition of African catfish. *African Journal of Agricultural Research* 8(37): 4604–4607.
- [38] Yaqub, H. B. (1999). Earthworm and maggot meals

as a potential fish meal replacement. Dissertation submitted to the Institute of Renewable natural Resources Farm, University of Science and Technology, Tema. 18pp.

- [39] Meena, K. D. (2015). Reguration and perspective of feed intake in fish. [http://www.aquafind.com/articals/ feed intake-in-fish.php] site visited on 05/05/2018.
- [40] Mohamed, E. H. A. and Al-Sabahi, G. N. (2011). Fatty acids content and profile of common commercial Nile fishes in Sudan, *International Journal of Fisheries and Aquaculture*, vol. 3, no. 6, pp. 99-104.
- [41] Mwanja, M., Nyende, D. L., Kagoda, S., Munguti, J. and Mwanja, W. W. (2010). Characterisation of fish oils of mukene (*Rastrineobola argentea*) of Nile basin waters-Lake Victoria, Lake Kyoga and the Victoria Nile River. *Tropical Freshwater Biology* 19: 49-58.
- [42] Henderson, R. J. (1996). Fatty acid metabolism in freshwater fish with particular reference to polyun-saturated fatty acids. *Arch Tierernahr* 49(1): 5-22.
- [43] Bachok, Z., Mfilinge, P. and Tsuchiya, M. (2006). Characterization of fatty acid composition in healthy and bleached corals from Okinawa, Japan. *Coral Reefs* 25: 545–554.
- [44] Cintra, D. E., Ropelle, E. R., Moraes, J. C., Pauli, J. R. and Morari, J. (2012). Unsaturated Fatty Acids Revert Diet-Induced Hypothalamic Inflammation in Obesity. *Plos One* 7(1): 1-20.
- [45] Zenebe, T., Ahlgren, G. B., Gustafsson, I. and Boberg, M. (1998). Fatty acid and lipid content of *Oreochromis niloticus* in Ethiopian lakes – Dietary effects of phytoplankton. *Ecology of Freshwater Fish* 7: 146-158.